

MetaXpress® 6 Software Guide

Acquiring a Plate with Timelapse and Fluidics Enabled

UNLEASH YOUR BRILLIANCE

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Chapter Purpose

The purpose of this chapter to guide the user through setting up a timelapse acquisition with fluidics events. This includes selecting objectives, plates, wavelengths, focal position, and configuring fluidics events.

The fluidics option has a single channel pipettor that can be used to add compounds, exchange media, and wash cells as part of an imaging experiment.





Fluidics Option: Hardware Components







Fluidics Option: 96-Well Plate Setup



Fluidics Option: 384-Well Plate Setup



Preparing for a Fluidics Experiment

Preparation

- 1. Turn on the ImageXpress Micro system at least two hours before experiment to allow the system temperature to stabilize
- 2. Set sample and compound plate temperature on the options controller box
- 3. Make sure system has the appropriate objective(s) and filter sets
- 4. Place the EC sealing ring in the system
- 5. (optional) Put your plate in the system

After system has equilibrated

- Check status of environmental control (temperature/CO₂/ Humidity). In the main menu, select Control > ImageXpress > Environment Control
- 2. Load tips and compound plate(s)
- 3. Load sample plate without lid if not already in the system
- 4. If using CO_2 / Humidity:
 - Place EC sealing ring sample plate inside system
 - Check gas tank and regulator
 - Check liquid level in water reservoir



1 Environment Control	- • •
Current Temperature:	37.0 C
Temperature Setpoint:	37.0 C
Compound Plate Temperature:	37.0 C
Compound Plate Temperature Setpoint:	37.0 C
CO2 Pressure:	OK
CO2 Pump State:	On
Liquid Level:	ОК
	Close

NOTE: Set Compound and Sample Plate Temperatures on the options controller box. Outof-focus images may result if plates are not equilibrated. If plates need to be equilibrated, place plates in system with sealing ring for at least 20 minutes prior to experiment.





- 1. Open Plate Acquisition Setup
 - In the main toolbar click on



OR

- Under the Screening menu, select Plate Acquisition Setup
- 2. Click on the **Configure** tab





- 3. Select the **Objective and Camera** tab
- 4. Select the desired **Magnification** from the drop-down menu
 - You may need to adjust the correction collar of the objective; refer to the Main Taskbar to do this.
 - Select **Camera Binning** (please see next slides for guidance)
 - Pixel size is automatically calculated based on magnification and binning
 - Set Camera Binning to 1 to acquire unbinned images maximum resolution
- 5. If the Gain option appears, start with gain set to Low

Objective and Camera- 4X S Flu				-	
Plate- Corning 1536-well Black-	Magnification:	4X S Fluor	-		
Sites to Visit- multi-well	Comoro binnina:	1	Calibration (binned):	1.61 × 1.61 um	
Acquisition	Camera birming.		Calibration (binned).	1.01X 1.01 UIII	
Autofocus	Gain	Low -			
Wavelengths	Cidii 1.	Low			
W1 DAPI					
W2 FITC					
Display					
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What is Binning?

Combining groups of pixels into a single pixel during image acquisition



Example of 2x2 Binning

Each pixel records an intensity







Image

4 Pixels are summed to make one larger pixel





Why Bin?

Brighter pixels

• The resultant pixel is brighter than any of the 4 component pixels

Save Space

• 2x2 binning reduces file size 4-fold

Increase Speed

- Faster image transfer from MetaXpress to database
- Faster image analysis

When to Bin

- You do not need to see intricate sub-cellular detail
- Cell counting
- Scoring cells positive or negative for fluorescent markers
- Measuring overall cell intensity





- 6. Select the Plate tab
- 7. Select the appropriate Plate Type from the drop-down menu







- 8. In the Plate Section, select the wells you would like to acquire
 - Left click and drag mouse to select wells
 - Click on "All" (top left corner), row letters, column numbers, or individual wells
 - Gray wells are deactivated, green wells are activated and will be imaged
 - Right click on a well to move the stage to that position (well turns dark green)

NOTE For experiments involving fluidics, select all wells to be acquired. Well selection for individual fluidics events will be done separately.







9. Click on the Sites to Visit tab

- Select **Single Site** to acquire one site in the middle of the well (recommended for fast kinetic experiments)
- To acquire a single site elsewhere in the well, refer to the next section on setting up multiple sites

Objective and Camera- 10X Plar Plate- Greiner 384-well thin bot: Sites to Visit- single site Acquisition	Site Options Single site Fixed number of sites Adaptive acquisition Multi-well	Custom field of v X: 50 + Y Site/image size: 1.3	riew (%): ☆ 50 🗼 9 x 1.39 mm	Well size: 11 mm ² Number of sites: 1 17.82% Well Coverage	
Autofocus Wavelengths W1 DAPI W2 FITC	Acquires a single site ce	entered in each well			
Display					





9. On the Sites to Visit tab

- Select **Fixed number of sites** to acquire multiple sites
- Build site grid by specifying number of Columns and Rows
- Spacing defines the x-y spacing between sites

Objective and Camera- 10X Plan	Site Options	Custom field of view (%):	Well size: 11 mm ²
Plate- Greiner 384-well thin bot:	 Single site 	×: 50 🜩 Y: 50 🜩	Number of sites: 2
Sites to Visit- multi-site	Adaptive acquisition	Ste (mage size: 1.29 x 1.29 mm	35.65% Well Coverage
Acquisition	Multi-well	Site/indge size: 1.35 X 1.35 min	
Autofocus	Acquires a fixed number	of sites in each well	
Wavelengths	_		
W1 DAPI	Con		
W2 FITC	Columns: 2 🖨 0	Tile sites	
Display	Rowe: 2 🌰 0	Et sites to well	
	Hows		
		Overlap sites 10%	

- **Tile sites** places sites edge to edge
- Fit sites to well spreads sites to well edge
- Overlap sites 10% overlaps edges of sites for stitching







NOTE Left clicking on site selects (green) or deselects (gray) for imaging. Right click moves stage to that position (dark green)

- 9. On the Sites to Visit tab
 - The Adaptive acquisition and Multi-well options are not compatible with fluidics events

Objective and Camera- 10X Plar Plate- Greiner 384-well thin bot: Sites to Visit- single site Acquisition	Site Options Single site Fixed number of sites Adaptive acquisition	Custom field of view (%): X: 50 Y: 50 Site/image size: 1.39 x 1.39 mm	Well size: 11 mm ² Number of sites: 1 17.82% Well Coverage
Autofocus	Acquires a single site ce	ntered in each well	
Wavelengths			
W1 DAPI			
W2 FITC			
Display			





- 10. Click on the Acquisition tab
 - Always Enable laser-based focusing
 - Enable Acquire Time Series
 - Enable Use Fluidics
 - Optionally, enable Perform shading correction

Objective and Camera- 10X PF	Autofocus options
Plate- 384 Wells (16x24)	Enable laser-based focusing
Sites to Visit- single site	Enable image-based focusing (for acquisition or laser recovery)
Acquisition	Acquisition options
Autofocus	Acquire Time Series
Wavelengths	Acquire Z Series
W1 DAPI	
W2 FITC	
Timelapse- 6 time points	Use Fluidics
Fluidics	Run Journals During Acquisition
Display	Analyze Images After Acquisition
	Perform shading correction Directory C:\





11. Click on the Timelapse tab

- From the drop-down menu, select an option to **Perform time series for**:
 - **One well then the next** for fast kinetic time lapse in a single well (or site). All time points will be collected in one well before moving to the next well.
 - All selected wells for long-term time lapse experiments. All wells are acquired during each time point.
 - One row then the next or One column then the next for specialized cases (uncommon for fluidics experiments).

Objective and Camera- 10X PF		21	
Plate- 384 Wells (16x24)	Number of timepoints:	21	
Sites to Visit- single site	Perform time series for:	One well then the next 🔹	
Acquisition	Approximate minimum	One well then the next	
Autofocus	Interval:	One row then the next	
Wavelengths		All selected wells	
W1 DAPI	Duration:	20 🜩 sec 🔻	
W2 FITC			
Timelapse- 6 time points			
Fluidics			
Display			



11. On the **Timelapse** tab

- Enter the Number of timepoints
- Set the Interval: time between each image taken (ms, sec, min, or hr)
- Set the **Duration**: total time of experiment (ms, sec, min, or hr). This is equivalent to Interval x Number of timepoints

Objective and Camera- 10X PF	Number of the second states	21
Plate- 384 Wells (16x24)	Number of timepoints:	21
Sites to Visit- single site	Perform time series for:	One well then the next 🔹
Acquisition	Approximate minimum	time interval: 150.0
Autofocus	Interval:	1
Wavelengths		
W1 DAPI	Duration:	20 🌲 sec 🔻
W2 FITC		
Timelapse- 6 time points		
Fluidics		
Display		





- 12. Click on the **Fluidics** tab to:
 - **Configure Stations**: configure the deck layout, select tips and compound plate type, set global pipetting parameters
 - Add New Event: schedule fluidics events
 - **Reset tips**: reset tip and liquid levels

Objective and Camera- 10X PF		Configure Stations
Plate- 384 Wells (16x24)	Scheduled Events:	Configure Stations
Sites to Visit- single site	Time Event	
Acquisition		
Autofocus		
Wavelengths		
W1 DAPI		
W2 FITC		
Timelapse- 6 time points		
Fluidics		
Display		
	Reset Tips Add new Event Delet	e Event Edit Event





Configuring Stations

On the Fluidics tab, click on the Configure Stations button





Tracking Liquid Levels

In the **Configure Fluidic Stations** dialog Molecular Devices recommends enabling **Track Volume** and **Track Liquid Surface**

- Enabling **Track Volume** will track available volume and allow the software to warn you if there is not enough volume for scheduled fluidic events
- Enter the appropriate values for Initial Volume for Tracking in compound plates and sample plate
- Enabling **Track Liquid Surface** allows the system to draw or dispense liquid at the surface and move the tip up or down as the liquid level changes. This requires enabling **Track Volume**, an accurate plate template, and correct **Initial Volume** settings





NOTE If **Track Liquid Surface** is not checked, the pipet tip will be inserted near the bottom of the well for draw and dispense events



Configuring System Properties

- In the Configure Fluidic Stations dialog, click on System Properties button
- 2. Click on the **Help** button for explanations on each property
- 3. Items to note:
 - Draw Rate and Dispense Rate can affect:
 - Length of experiment, volume accuracy, disturbance to cells
 - Post Draw Transport Air Gap minimizes fluid dripping
 - Pre Draw Air Gap and Smart Dispense help push fluid out
 - When Wet Dispense is enabled
 - Tip is just below liquid surface in sample plate when Track Liquid Surface is on
 - Tip is just above well bottom in sample plate when **Track Liquid Surface** is off
 - When Wet Dispense is disabled
 - Tip is just above the liquid in sample plate
 - Draw Overfill draws extra compound for better volume accuracy
 - Molecular Devices suggests using the Pump Settle Time default value



	Fluidics System Properties		x
	Draw Rate (ul/s):	25	-
	Dispense Rate (ul/s):	250	-
	Post Draw Transport Air Gap (ul):	2	-
1	Pre Draw Air Gap (ul):	10	-
	Wet Dispense:		
	Smart Dispense (ul):	1	×
	Draw Overfill (ul):	0	-
	Pump Settle Time (ms):	500	-
	OK Cancel	Help	

NOTE the settings in **Fluidics System Properties** are saved with the protocol and used for all fluidics events



Optimizing System Properties

Molecular Devices highly recommends testing and optimizing fluidics settings for any experiment. As the example below shows, multiple properties can affect results. | Fluidics Event







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Time

Event Scheduling

- 12. On the **Fluidics** tab, click on the **Add New Event** button to begin setting up a fluidics event
 - Events can be scheduled to occur before or after any time point (see diagram below)
 - If time points are close together, a fluidics event may delay acquisition of subsequent time points









DEVICE

Event Scheduling – Compound Addition Event

Compound addition adds reagent to the sample plate at the designated **Time point**:

- i. Imaging is paused
- ii. Pipettor loads a tip
- iii. Aspirates reagent from compound plate
- iv. Adds reagent to the corresponding well in sample plate (and optionally mixes)
- v. Disposes tip
- vi. Imaging continues

To configure this dialog:

- Enter the **Volume** to be drawn from the compound plate and dispensed into the sample plate
- Optionally select Number of Mixes and Mix Volume
- Enter a value for **Mix Dead Volume** volume that remains in tip during mix to prevent bubbles and is dispensed out at the end of mix cycle
- Optionally enable Mix while imaging
- Select wells to apply these parameters to

Fluidic Event	×
Time point: 5	Before imaging
Event Type:	
Compound addition	
Washout	
 Journal 	provide the second second second
Compound plate:	Plate 1
Tip:	96, 200ul, FLIPR 🔻
Volume (ul):	10
Number of Mixes:	0
Mix Volume:	40 🚔 🔲 Mix while imaging
Mix Dead Volume:	10
Wells Affected:	
Al wells	
Selected wells	Select Wells
OK Test at A	.1 Cancel

NOTE REagent drawn from the compound plate is dispensed to the corresponding well in the sample plate. For example, draw from compound plate A01 and dispense to sample plate A01.





Event Scheduling – Wash Out Event

A wash out event performs a wash (or media refresh) in the sample plate. At the designated **Time point**:

- i. Imaging is paused
- ii. Pipettor loads a tip and aspirates liquid from sample plate
- iii. Disposes liquid (and optionally tip)
- iv. Aspirates liquid (new tip optional) from corresponding well of compound plate
- v. Dispenses liquid into sample plate
- vi. Repeats ii v as specified in Number of exchanges
- vii. Disposes of tip
- viii. Imaging continues

To configure this dialog:

- Enter the **Volume** to aspirate and dispense
- Specify the Number of exchanges
- Optionally Enable New tip for each exchange
- Select wells to apply these parameters to

Plate 1
96, 200ul, FLIPR -
10 🗢
3 🗘 🕅 New tip each exchange

NOTE Reagent drawn from the compound plate is dispensed to the corresponding well in the sample plate. For example, draw from compound plate A01 and dispense to sample plate A01.







Event Scheduling – Journal Event

A journal event runs a journal (macro) at the specified time point. A journal:

- i. Is a script written to customize fluidics steps in a protocol
- ii. Can be used to serial dispense compound from one well into multiple sample wells
- iii. Can be used to save media from sample plate for later analysis
- iv. Can be used for staggering pipetting events
- v. And more

To configure this dialog:

- Select Journal under the Event Type section
- Click on the **Open File** icon to navigate to the location of the previously configured journal
- Select wells to apply these parameters to







Reset Tip and Liquid Levels

- 12. On the Fluidics tab, click on the Reset Tip button to
 - Reset tips after loading a full tip rack (system keeps track of tips used during testing and acquisition)
 - Reset liquid levels after loading fresh compound plates
 - Reset liquid level after inserting a fresh sample (imaging) plate

NOTE A reminder to reset tips and/or liquid levels will always appear when you press the **Acquire Plate** button

Reset Tips / Liquid Levels	×
Reset tips	
Reset Tip tray 1: 96, 200ul, FLIPR Pipette Tips now present: 96	
Reset Tip tray 2: 96, 200ul, FLIPR Pipette Tips now present: 96	
Reset liquid levels	
Reset Compound plate 1	
Reset Compound plate 2	
Reset Sample plate	
	Close





13. Click on the Autofocus tab

- Select the appropriate option for **Well to well autofocus**:
 - Focus on well bottom: most scenarios using 10X and higher objective
 - Focus on plate bottom then offset by bottom thickness: for low magnification objectives (2X, 4X), thin plates, or microscope slide/coverslip.
 - Focus on plate and well bottom: for warped plates (plate bottom variation is more than half the optical thickness)

Objective and Camera- 10X PF	Laser-based Focusing
Plate- 384 Wells (16x24)	Configure Laser Settings
Sites to Visit- single site	Well to well autofocus Focus on well bottom
Autofocus	Image-based Focusing Focus on plate bottom, then offset by bottom thickness
Wavelengths	Algorithm: Standard - Focus on plate and well bottom
W1 DAPI	
W2 FITC	Allow image-based focusing for recovery from laser-based well bottom failures
Timelapse- 6 time points	
Fluidics	
Display	Initial well for finding sample First well acquired
	Number of wells to attempt initial find sample 3
	Timelapse Autofocus First timepoint only
	View Focusing Details





13. On the Autofocus tab

- Set Initial well for finding sample to First well acquired
 - This serves as a check to verify a plate is loaded
 - Only disable for very specific applications (i.e., oil immersion objectives)
- Set Number of wells to attempt initial find sample to 3

Objective and Camera- 10X PF	Laser-based Focusing
Plate- 384 Wells (16x24)	Configure Laser Settings
Sites to Visit- single site	
Acquisition	Well to well autorocus Focus on well bottom
Autofocus	Image-based Focusing
Wavelengths	Algorithm: Standard Binning: 2 A Custom exposure times
W1 DAPI	
W2 FITC	Allow image-based focusing for recovery from laser-based well bottom failures
Timelapse- 6 time points	
Fluidics	
Display	Initial well for finding sample First well acquired
	Number of wells to attempt initial find sample 3
	Timelapse Autofocus





13. On the Autofocus tab

- Select the appropriate option for **Timelapse Autofocus** from the drop down menu:
 - **First timepoint only** for fast kinetic time lapse in a single well (i.e., the stage does not move between time points). Use this setting when selecting One well then the next on the Timelapse tab.
 - All timepoints for long-term time lapse where the stage moves from well to well between time points. Use this setting when selecting All selected wells on the Timelapse tab.
 - **Every Nth timepoint** for slower or longer kinetic experiments in a single well to periodically verify focal position. This setting is recommended when selecting One well then the next on the Timelapse tab.

Display	Initial well for finding samp	le First well acquired	• A	▼ 1 [↑] / _▼
	Number of wells to attemp	t initial find sample 1		
	Site Autofocus	All sites	•	
	Timelapse Autofocus	First timepoint only	v 2	×.
		First timepoint only		
		All timepoints Every Nth timepoint		



- 14. Select the Wavelengths tab
 - Select the number of wavelengths or channels that you will acquire on this plate
 - A separate **W** tab will appear below for each channel
 - You can select up to 8 wavelengths

Objective and Camera- 10X Plar		2	
Plate- Greiner 384-well thin bot:	Number of wavelengths:	2	
Sites to Visit- multi-site			
Acquisition			
Autofocus			
Wavelengths			
W1 DAPI			
W2 FITC			
Display			





15. Select the W1 tab

- Select correct **Illumination Setting** from the drop-down menu
- In the Plate Map, right-click to select a site/well that should contain the highest signal for the wavelength chosen

Objective and Camera- 10X Plan Plate- Greiner 384-well thin bot:	Illumination setting: DAPI
Sites to Visit- multi-site	Exposure (ms): 50 - Auto Expose Target max intensity: 33000
Acquisition	Autofocus options
Autofocus	Part lager
Wavelengths	offset (um)
W1 DAPI	Laser with z-offset 12.36
W2 FITC	
Display	
	Calculate Offset ✓ Use Z stack □ Custom Range Range (um) Step (um) 138.89 ↓ 5.56 ↓





15. On the W1 tab

- Click on the **Calculate offset** button to perform an automatic routine for finding the best focal position (Post-laser offset value)
 - Enable **Use Z Stack** for an interactive option to select the focus position. The software will acquire a Z stack of images and allow you to select the most in-focus image.
 - Enable **Custom Range** to specify a custom range and step size for the focus search

Objective and Camera- 10X Plar Plate- Greiner 384-well thin bot:	Illumination setting: DAPI
Sites to Visit- multi-site	Exposure (ms): 50 🖨 Auto Expose Target max intensity: 33000
Acquisition	
Autofocus	Part lagar
Wavelengths	offset (um)
W1 DAPI	Laser with z-offset 12.36
W2 FITC	
Display	
	Calculate Offset Vise Z stack Custom Range Range (um) Step (um) 138.89 5.56
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What is a Post- Laser Offset?

Post-laser offset is the Z distance between the bottom of the well and the sample

- Laser autofocus routine finds the well bottom, NOT the biological sample of interest
- You may need to empirically determine the offset (or distance) between the well bottom and the sample
- Very wavelength dependent (chromatic aberration)
- Offset can be positive or negative
- It is recommend to checking multiple wells for consistency





15. On the W1 tab

- Enter an Exposure (ms) time and click the Focus button
 - Evaluate the image for pixel intensity range (bit range)
 - For fluidics experiments you may need to decrease/increase exposure time according to expected results. For example, if an increase in intensity is expected after the fluidics event, set the exposure so that the first time point is between 10-25% of the bit range of the camera. It may be necessary to run test wells in order to determine the optimal exposure time for the entire experiment.

Objective and Camera- 10X Plar	Illumination setting: DAPI
Plate- Greiner 384-well thin bot:	
Sites to Visit- multi-site	Exposure (ms): 50 🖨 Auto Expose Target max intensity: 33000 🖨
Acquisition	
Autofocus	Part lager
Wavelengths	offset (um)
W1 DAPI	Laser with z-offset
W2 FITC	
Display	
	Calculate Offset ✓
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15. On the W1 tab

- Select the appropriate option from the Acquisition Options drop-down menu to specify how often to collect the selected wavelength
 - At all time points acquires this wavelength at each time point
 - At start of experiment acquires this wavelength only at time point 1
 - At start/end of experiment acquires this wavelength at only the first and last time points
 - Every nth time point acquires this wavelength every nth time point (2nd, 5th, 6th, etc.) throughout the time lapse experiment

NOTE This option is only available when your acquisition protocol contains more than one wavelength.

W1 DAPI W2 FITC	Laser with z-offset	
Timelapse- 2 time points		
Display	Calculate Offset Image Use Z stack Custom Range Range (um) Step (um) Acquisition Options Acquisition Options	
	Timelapse: at all time points	
38	at all time points at start of experiment at start/end of experiment every nth timepoint	ULAR

- 16. Click on the **W2** tab (and subsequent W tabs)
 - Select desired Illumination Setting from the drop-down menu
 - In the Plate Map, right-click to select a site/well that should contain the highest signal for the wavelength chosen
 - Calculate Focus offset
 - Determine Exposure time
 - Select appropriate Acquisition Options for Timelapse

Objective and Camera- 10X Plan	
Plate- Greiner 384-well thin bot:	Illumination setting: FIIC
Sites to Visit- multi-site	Exposure (ms): 100 🖨 Auto Expose Target max intensity: 33000 🚔
Acquisition	Autofocus options
Wavelengths	Offset (um)
W1 DAPI	Z-offset from W1 2.76
W2 FITC	
Timelapse- 2 time points	
Display	Calculate Offset Vise Z stack Custom Range Range (um) Step (um) 138.89 5.56
	Acquisition Options
	Timelapse: at all time points





- 17. Click on the **Display** tab to configure:
 - Auto Arrange Images: Software automatically determines the arrangement and size of images shown during acquisition.
 - Click on Display Acquisition Layout: Manually configure how the images will look during acquisition (position, size, scaling, monochrome or color).
 - **Display images during autofocus** should be checked to help with finding post-laser offset.
 - **Display images during acquisition** displays images according to the settings determined using Auto Arrange Images or Display Acquisition Layout.
 - **Display a color overlay of wavelength images during acquisition**: Will create a color composite of the first 3 wavelengths selected.

Objective and Camera- 10X Plar		
Plate- Greiner 384-well thin bot:		
Sites to Visit- multi-site	Auto Arrange Images	
Acquisition	Display, Acquisition Layout	
Autofocus	Display Acquisition Layout	
Wavelengths	Display images during autofocus	
W1 DAPI		
W2 FITC		
Display	Display a color overlay of wavelength images during acquisition	



- 18. Click on the **Save Protocol** button at the bottom of the Plate Acquisition Setup dialog
 - A star on the Save Protocol button indicates there are unsaved changes to the protocol
 - It is recommended to save your settings to a file, rather the database
 - Click on **Save**, name the protocol, and navigate through windows to save the file (.hts)





19. Click on the Run tab and enter:

- Folder Name: folder your plates go in in the database (i.e. project or PI)
- **Plate Name**: the name of the plate to be imaged (i.e. specific experiment)
- Barcode (optional): manually enter the plate barcode
- **Storage Location**: select where you want images to be stored (there may only be one choice)
- **Description**: enter any identifying information you would like to store with the plate

Configure Run	Active Wavelength	FITC	•	Snap Start Live	60 Focus	Test	Preview
Folder Name	Transfluor	Barcode					
Plate Name	Transfluor 10x	Description	Transfluor plate		*		
Storage Location	Local File Server				~	Acquire Plate	
	Exposure Time (ms)	Snap	Test	Focus Offset (µm)			
DAPI	Auto Expose 50 🖨	[` O "		Calculate 12.36	•		
FITC	Auto Expose 400 🖨	[`0"		Calculate 2.76	•		
42						MOL	ECULAR

20. Click on the **Acquire Plate** button to begin acquisition of the plate

Configure Run	Active Wavelength	FITC	•	Snap Start Live	Focus	Test	Preview
Folder Name	Transfluor	Barcode					
Plate Name	Transfluor 10x	Description	Transfluor plate		*		
Storage Location	Local File Server				Ψ.	Acquire Plate	
	Exposure Time (ms)	Snap	Test	Focus Offset (µm)			
DAPI	Auto Expose 50 🚔	[`O"		Calculate 12.36	▲ ▼		
FITC	Auto Expose 400 🚔	[°O]		Calculate 2.76	* *		





Support Resources

- F1 / HELP within MetaXpress® Software
- Support and Knowledge Base: <u>http://mdc.custhelp.com/</u>
- User Forum: <u>http://metamorph.moleculardevices.com/forum/</u>
- Request Support: <u>http://mdc.custhelp.com/app/ask</u>
- Technical Support can also be reached by telephone:
 - 1 (800) 635-5577
 - Select options for Tech Support → Cellular Imaging Products → ImageXpress Instruments





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