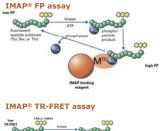
Use of IMAP[®] TR-FRET detection mode for the IMAP[®] AGC and CAMK Substrate Finder

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Abstract

One of the main bottlenecks in the development of in-vitro kinase assays is the identification of suitable peptide substrates for new kinases and their subsequent optimization for HTS performance. To this end, Molecular Devices has developed a number of Substrate Finder plates to run with its IMAP® platform. The IMAP® Technology utilizes the high affinity binding of phosphate to immobilized trivalent metals with two detection modes: TR-FRET and FP. We have previously described the equivalency of the assay results between TR-FRET and FP modes Finder plate. Here we show this equivalency for the AGC and CAMK group Substrate Finder plate.



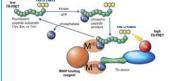


Figure1: Assay principle for IMAP® TR-FRET and IMAP® FP assay

Introduction

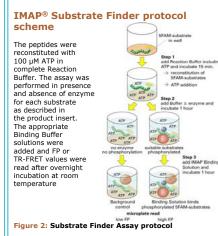
Identification of a suitable substrate for known or novel kinases has proven itself to be one of the bottlenccks in kinase screening assay development. Molecular Devices offers three Substrate Finders for Ser/Thr or Tyr kinases in their IMAP® platform which have proven themselves to be valuable tools to this end. With the introduction of the IMAP® TR-FRET detection, the need to also run these tools in TR-FRET detection, the need to also run these tools adapted the Ser/Thr 1 (CamK/AGC group) Substrate Finder Kit (R8131) to TR-FRET detection and compared FP and TR-FRET detection for Pim1 and PKA kinases.

Optimization of the Substrate Finder protocol for TR-FRET

Substrate Finder peptides have been grouped by Binding Solution requirements with the goal of minimizing the number of Binding Solutions needed per plate. For use of TR-FRET detection, we chose three TR-FRET Binding Buffer conditions per original FP condition and ran the plate against a high concentration of PKA. Based on these data we chose the TR-FRET Binding Buffer conditions to replace the FP Binding Buffer conditions indicated in bold in Table 1.

	FP conditions	TR-FRET conditions
1	100%A	80%A, 20%B 1:600
	1:400	70%A, 30%B 1:600
		60%A, 40%B 1:600
2	75%A, 25%B	50%A, 50%B 1:600
	1:600	40%A, 60%B 1:800
		30%A, 70%B 1:800
3	60%A, 40%B	50%A, 50%B 1:600
	1:600	40%A, 60%B 1:800
		30%A, 70%B 1:800

Table 1: Binding Buffer conditions tested to adapt the Substrate Finder to TR-FRET detection (chosen ones in bold). All TR-FRET Binding Buffer conditions included Tb Donor at a dilution of 1:400.



Substrate Finder Results for Pim1

The Substrate Finder for Ser/Thr kinases 1 (R8131) was originally developed for FP detection. Using the example of Pim1 we show here that similar hits for potential substrates are identified using either FP or TR-FRET detection mode after Binding Solution optimization as shown in Table 1.

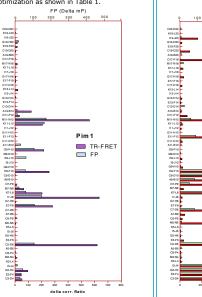


Figure 3: Pim1 (Upstate) was run at 0.5U/mL on the IMAP® Substrate Finder for Ser/Thr 1 (R8131) in the presence of 100µM ATP using Binding Buffers as described in Table 1 (bold for TR-FRET).

Position	Sequence	∆mP	TR-FRET △corr. Ratio
M11-N12	5FAM-AKRRRLSSLRA-OH	233	560
K11-L12	5FAM- ERMRPRKRQGSVRRRV-NH2	219	206
I7-J8	5FAM-RKRRQTSM-OH	200	633

sequence for FP and TR-FRET detection on Pim1.

Substrate Finder Results for PKA

As a second example for the fidelity of TR-FRET detection on the Substrate Finder for Ser/Thr kinases 1 (R8131), we chose PKA. Again the hits identified in FP detection mode match well with the hits identified in TR-FRET detection mode.

FP (Delta mP)

200 300

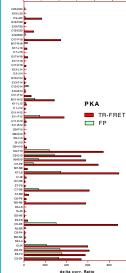


Figure 4: PKA (Upstate) was run at 0.5U/mL

on the IMAP[®] Substrate Finder for Ser/Thr 1 (R8131) in the presence of 100µM ATP using

Binding Buffers as described in Table 1 (bold

K7-L8 5FAM-GRTGRRNSI-NH2

C5-D6 5FAM-KKRPQRRYSNVF-OH 302

C3-D4 5FAM-LKKLTRRPSFSAQ-OH 325

Table 3: Substrate Finder hits position and

sequence for FP and TR-FRET detection on

for TR-FRET).

Positio

PKA

Further evaluation of hits from Substrate finder results in FP and TR-FRET

A: Pim1

Dilution curves for three of the positive Pim1 hits were run in FP (100 nM substrate)and TR-FRET detection mode (100 and 300 nM substrate). While the total EC_{30} did not change much between FP and TR-FRET detection, the latter resulted in lower standard deviations resulting in the option to run at lower enzyme concentrations. For all of the substrates a significant increase in assay window with increased substrate concentration could be observed as expected for TR-FRET.

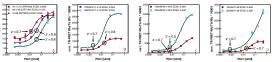


Figure 5: Dilution curves for selected hits from the Substrate Finder for Pim1 kinase. Circled data points indicate lowest concentration resulting in a z'-factor greater than 0.5. Panel A: FP detection, Panel B-D: TR-FRET detection.

B: PKA

Dilution curves for three of the positive PKA hits were run in FP and TR-FRET detection mode as described for Pim1 with similar results.

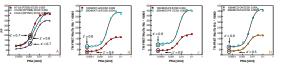


Figure 6: Dilution curves for selected hits from the Substrate Finder for PKA kinase. Circled data points indicate lowest concentration resulting in a z'-factor greater than 0.5. A: FP detection, Panel B-D: TR-FRET detection

Summary

TR-FRET

Ratio

890

877

536

AmP Acorr

345

- Identification of suitable kinase substrates is one of the major bottlenecks in Kinase assay development.
- The IMAP[®] Substrate Finders, totaling more than 150 peptides in three different plates, can shorten this process significantly.
- The IMAP® Technology offers two detection modes, FP and TR-FRET, both of which produce equivalent results as shown in this poster for the Ser/Thr 1 (CamK/AGC group) Substrate Finder kit (R8131).
- > The Substrate Finders are an integral part of the IMAP® platform for Kinase screening.

