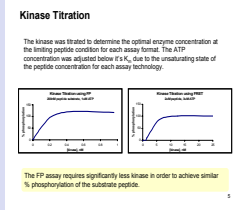
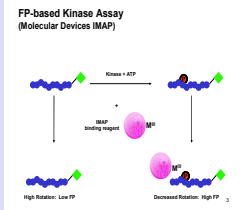




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Introduction

In recent years, a variety of non-radioactive assay technologies, such as Fluorescence Polarization (FP) and Fluorescence Resonance Energy Transfer (FRET), have been developed for HTS which offer the promise of increased throughput and cost savings. Two of these formats, the Invitrogen ZLYTE assay (FRET) and the Molecular Devices IMAP Progressive Binding System assay (FP), were evaluated as assay platforms for a serine kinase. An HTS assay was developed for each format using the appropriate substrate based on the same peptide sequence. The FP assay was selected for the HTS resulting in a robust campaign.



Comparison of HTS Conditions

	FP	FRET
[Substrate], μ M	0.200	2
[ATP], μ M	1	3
[Kinase], nM	0.030	2
% phosphorylation	18	60
Signal window	201	113
Z'	0.85	0.65
Signal:noise	18.3	12.4
Signal:background	5.8	3.4
Kinase run time, min	30	45
Signal development time	30	60
Plates processed in 24 hours	160	120

The FP assay was selected for the HTS campaign based upon reagent consumption and plate processing time.

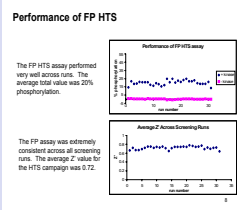
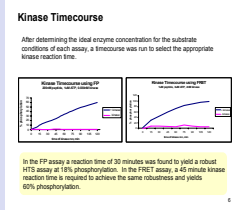
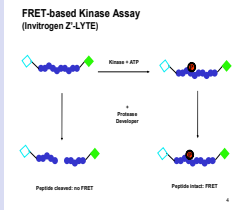
Conclusion

Two fluorescent assay technologies developed for kinases, the Invitrogen ZLYTE assay (FRET) and the Molecular Devices IMAP Progressive Binding System assay (FP), were evaluated for a serine kinase. Based on the comparison, we draw the following conclusions:

- An established peptide concentration for each assay technology, a robust FP assay can be run at 18% phosphorylation while the FRET assay must be run at >50% phosphorylation to achieve similar assay statistics.
- The FP assay used ~100 fold less kinase than required by the FRET assay.
- The FP HTS campaign showed consistency across all runs and was able to identify hits with a wide range of potency.

Format Comparison

Technology	IMAP	ZLYTE
Assay Format	FP (non-antibody)	FRET
Assay Type	Signal increasing	Signal decreasing
Detection Mode	Binding the phosphorylated peptide to IMAP maintains FRET signal	Phosphorylated peptide maintains FRET signal
Peptide Requirement	Sequence and labeling Limiting in assay to 200nM	Sequence modification and labeling Limiting in assay to 2 μ M
ATP	<100 μ M	μ M to mM



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